# SpatialData: an open and universal data framework for spatial omics

Quentin Blampey Luca Marconato

AI & ML in Spatial Single-Cell Transcriptomics Workshop 16 Oct 2025 - Lyon (France)







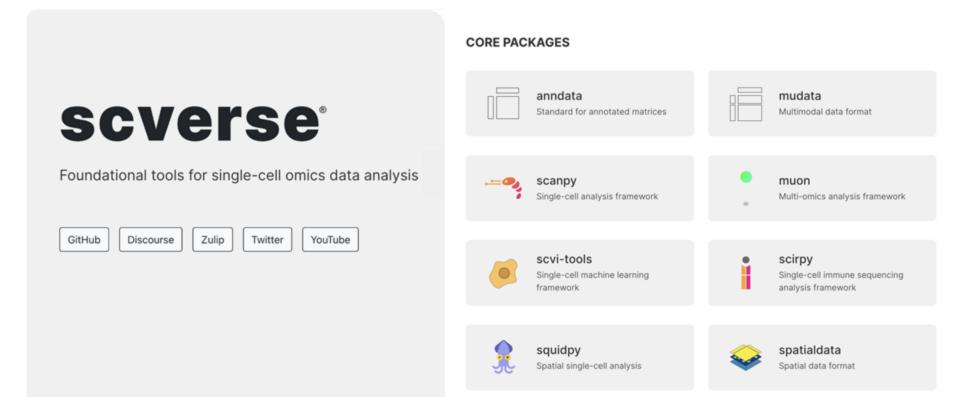
Documentation notebooks

Learning resources

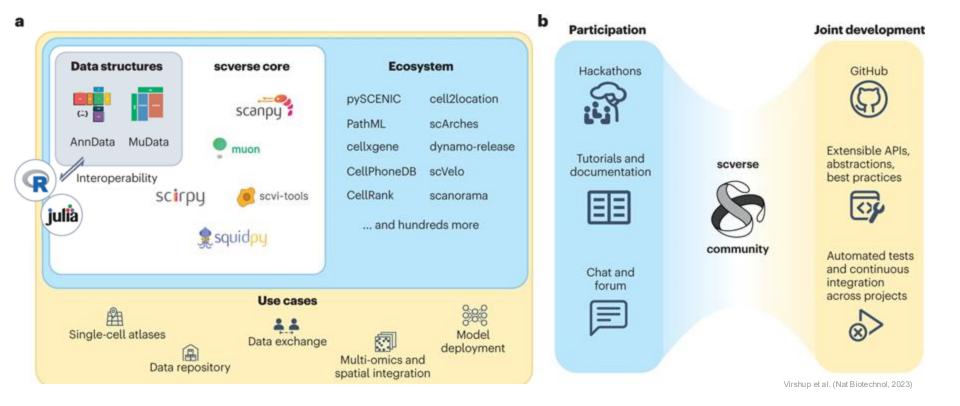




### We believe that interoperable foundational software can accelerate science



### We facilitate the growth of an ecosystem of methods, built upon our core libraries



#### Talks overview

#### From upstream packages to downstream tasks



Data structure for spatial omics

Official scverse data structure

As a contributor, I talk on behalf of scverse and Luca Marconato

nature methods



Analysis toolkit for spatial omics

scverse ecosystem package

Author and maintainer

nature communications



Deep learning / foundation model for spatial domains

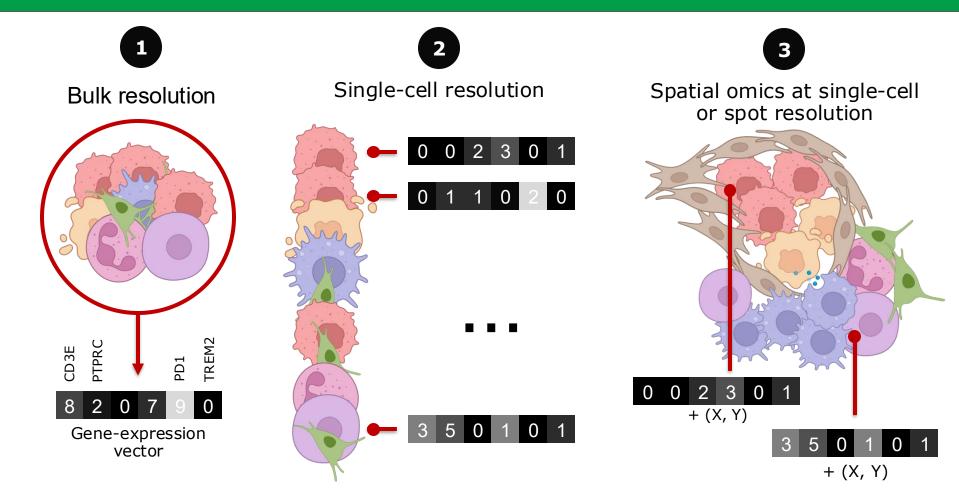
scverse ecosystem package

Author and maintainer

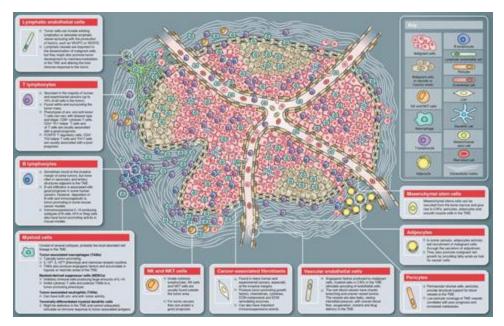
accepted in nature methods

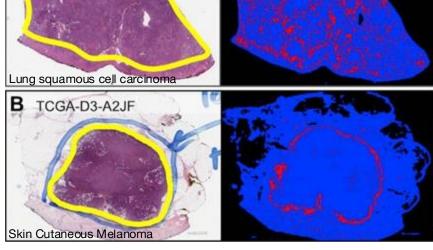
This talk

#### Spatial technologies enable the quantification of biological processes in the tissue context



# Spatial organization plays a crucial role in disease progression





TCGA-33-AASL

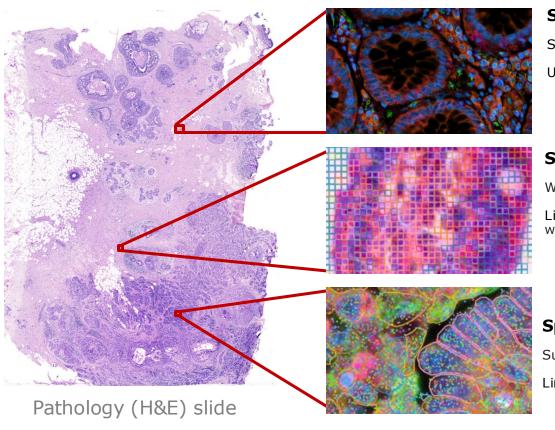
Adapted from Balkwill et al. (J. Cell Sci, 2018)

Adapted from Saltz et al. (Cell Reports, 2018)

Tumor microenvironments are organized in complex spatial structures

In this study, lymphocytes (red) infiltrating beyond the tumor boundaries (yellow) are linked with better prognosis

# Different types of omics data



#### **Spatial proteomics**

Staining images (one channel = one protein)
Up to 100 proteins (3 shown here)

#### **Spatial Transcriptomics (NGS-based)**

Whole-transcriptome sequencing

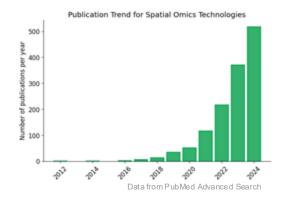
Limited to the spot resolution (subcellular resolution with the Visium HD)

#### **Spatial Transcriptomics (imaging-based)**

Subcellular resolution (3D transcript location)

Limited in number of different genes

#### Representing and operating on spatial omics data is becoming increasingly complex



Formats for

BaristaSea MIBI-TOF Cartana IIS Molecular Cartography DBiT-sea NanoString CosMx SMI **FISSEO** NanoString GeoMx DSP G4X Nova-ST **HDST** osmFISH IMC/Hyperion Open-ST LCM PLISH. MALDI Pixel-sea **MERFISH** Rebus Bio

RNA SPOTS RNAscope SABER-FISH seqFISH+ Seq-scope Slide-DNA-seq Slide-seq v2 smFISH SpaceM ST

XYZeq

Stereo-seq
StarMAP
Tomo-seq
Visium (SpaceRanger 1.0.0, 1.1.0, 1.2.0, 1.3.0, 1.3.1, 2.0.0, 2.0.1, 2.1.0, ...)

Visium HD
Xenium (Xenium Analyzer 1.0.2, 1.3.0, 1.4.0, 1.5.0, 1.6.0, 1.7.0, 2.0.0, 3.0.0, 3.0.1, ...)

Formats for raster data

ng jpg

Common compressed raster formats

.h5 .zarr

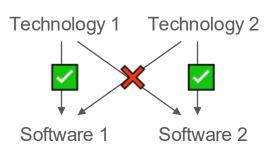
Tensor storages

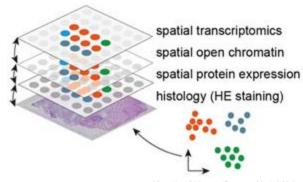


Multiscale storages



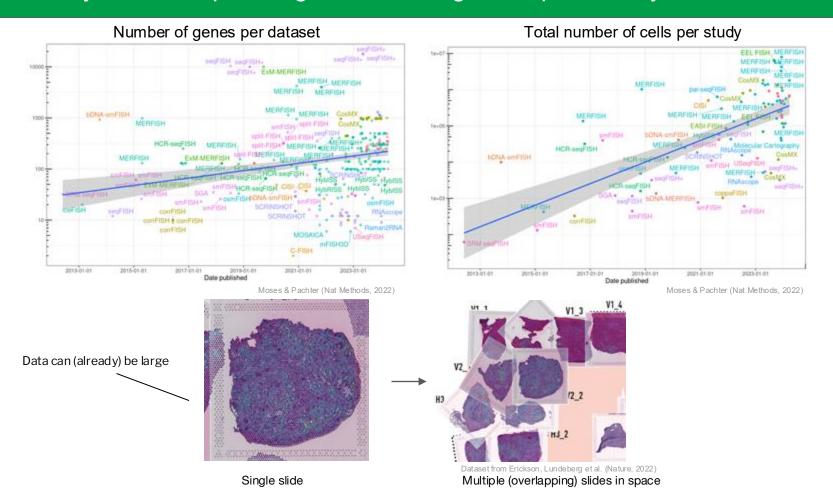
Formats for





Kiessling & Kuppe (Genome Med, 2024)

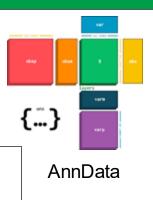
### Scalability becomes pressing as datasets grow exponentially in size



#### To reach the goal of building a foundational infrastructure, we bridged existing communities

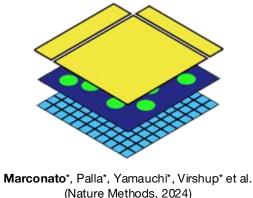


### Single-cell data





SpatialData



OME (Open Microscopy Environment)



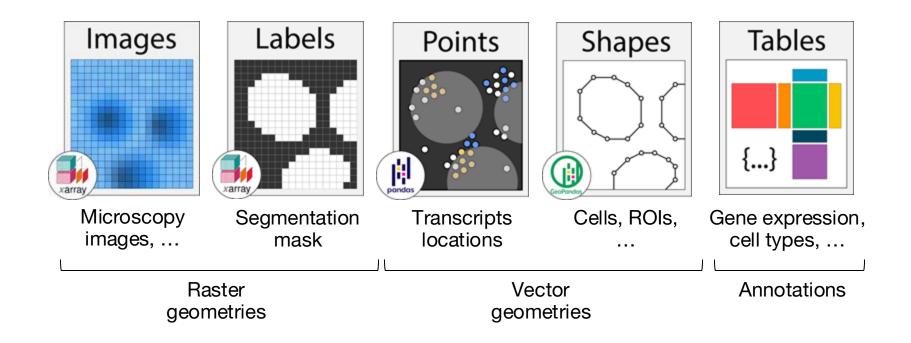
Large images, standard formats



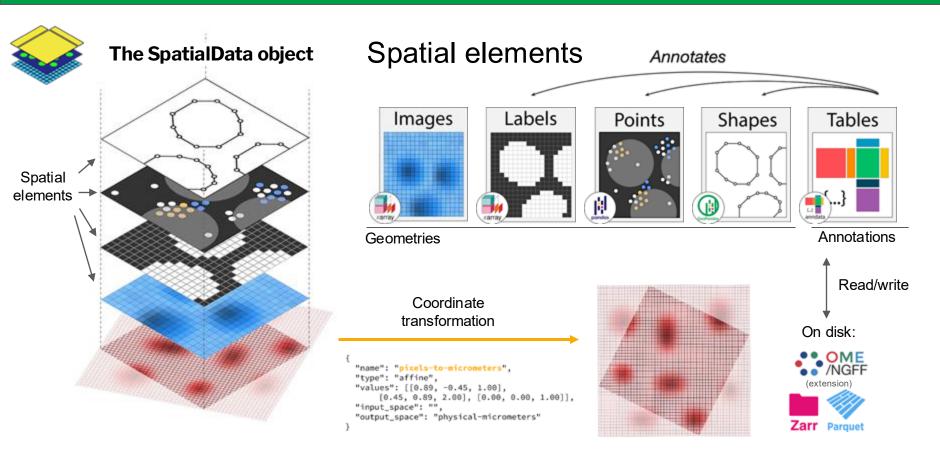
Napari core

napari
Interactive visualization

### Data is represented with a modular combination of reusable elements

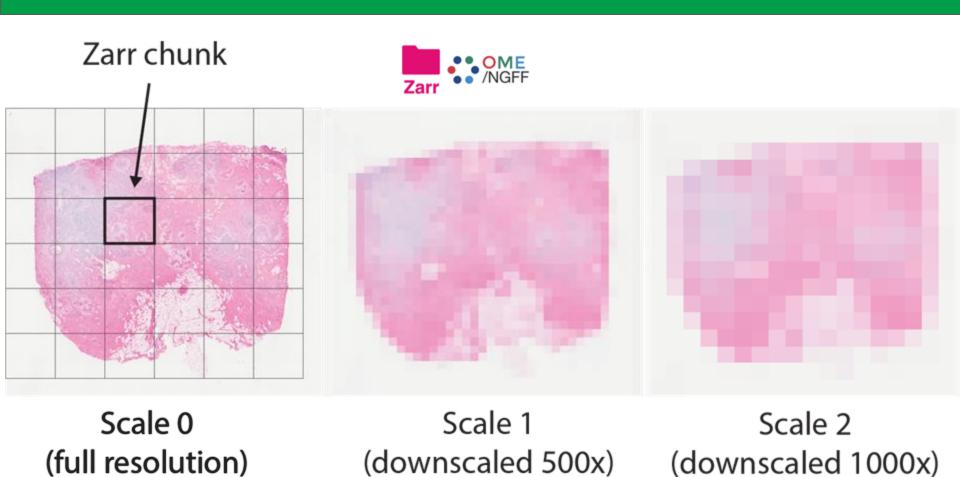


## SpatialData introduces a modular representation for spatial omics



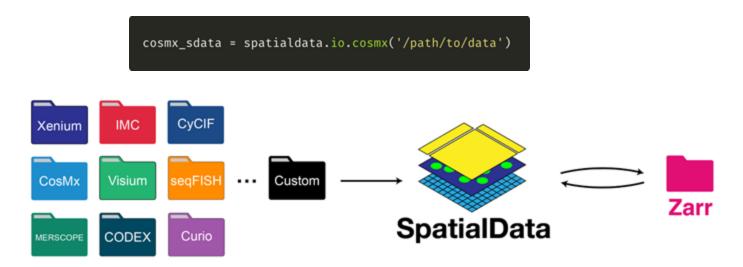
L. Marconato, O. Stegle, et al. (Nature Methods, 2024)

# Chunked, multiscale representation leads to efficient image access



### SpatialData IO streamlines data ingestion from popular technologies

### **SpatialData IO** (=Input Ouput)









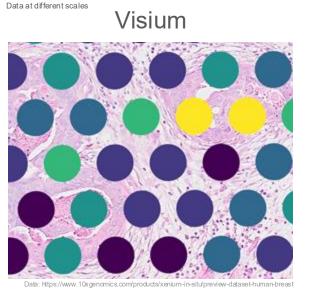
Laurens Lehner



Blampey



Quentin Wouter-Michiel Vierdag



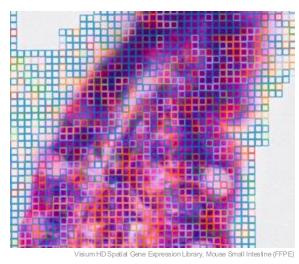
Resolution: 55µm Transcriptome-wide

#### Xenium



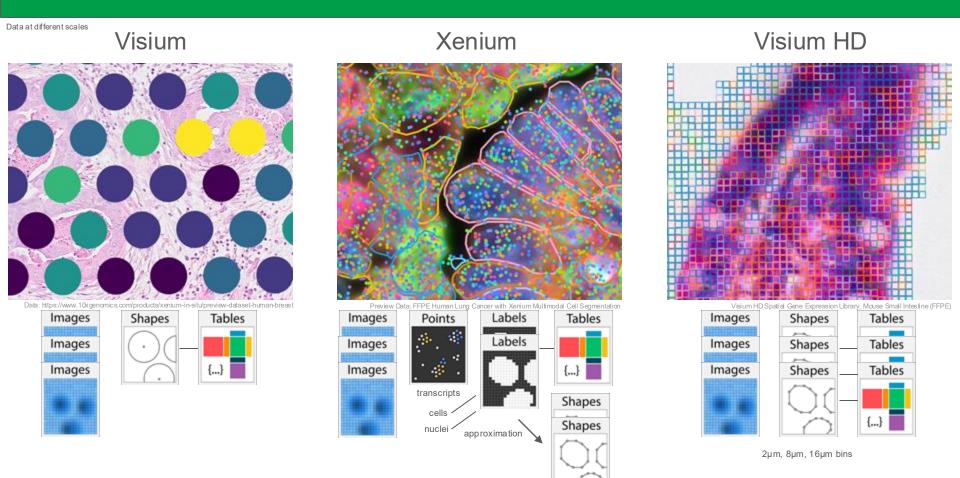
Resolution: single-molecule Up to 5K genes

#### Visium HD



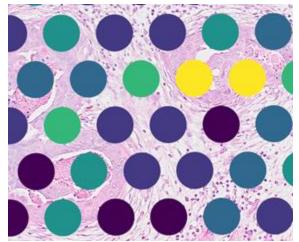
Resolution: 2µm, 8µm, 16µm, ... Transcriptome-wide

The raw data is very different across technologies



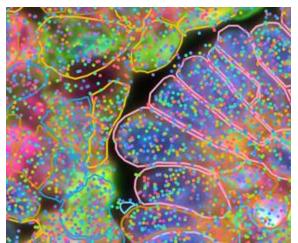
Data at different scales

#### Visium



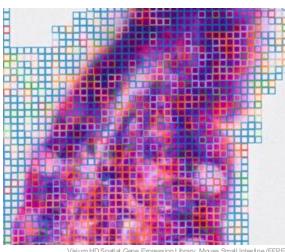
#### Data: https://www.10xgenomics.com/products/xenium-in-situ/preview-dataset-human-breast

#### Xenium



Preview Data: FFPE Human Lung Cancer with Xenium Multimodal Cell Segmentation

#### Visium HD



Visium HD Spatial Gene Expression Library, Mouse Small Intestine (FFPE)

#### Reading the data from disk:

```
from spatialdata_io import visium
sdata = visium('my_space_ranger_output')
```



```
from spatialdata_io import xenium
sdata = xenium('my_xenium_analyzer_output')
```



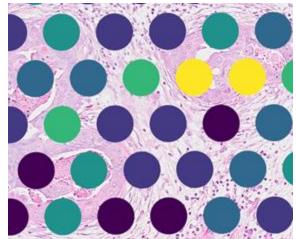




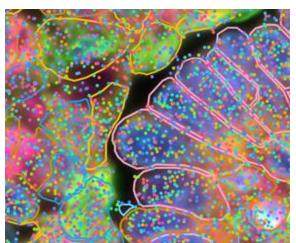
Xenium

Data at different scales

Visium

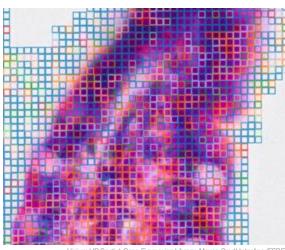


Data: https://www.10xgenomics.com/products/xenium-in-situ/preview-dataset-human-breast



Preview Data: FFPE Human Lung Cancer with Xenium Multimodal Cell Segmentation





Visium HD Spatial Gene Expression Library, Mouse Small Intestine (FFPE)

ion\_Muman\_Glioblastoma (Shapes)
 'downscaled\_lowres', with elements:
 (YtAssist\_FFPE\_Protein\_Expression\_Human\_Glioblastoma\_lowres\_image (Images), CytAssist\_FFPE\_Protein\_Expression\_Human\_Glioblastoma\_lowres\_image

CytAssist\_FFPE\_Protein\_Expression\_Human\_Glioblastoma\_lowres\_image (Images), CytAssist\_FFPE\_Protein\_Expr\_Human\_Glioblastoma (Shapes)

CytAssist\_FFPE\_Protein\_Expression\_Human\_Glioblastoma (Shapes)

print(sdata)

```
SpatialData object

Image:

- Image:
- "he_image": DataTree(cyx] (3, 5636, 1448), (3, 2818, 724), (3, 1469, 362), (3, 784, 181), (3, 352, 98)
- "he_image": DataTree(cyx] (1, 17698, 51187), (1, 8549, 25593), (1, 4274, 12766), (1, 2137, 639
8), (1, 1668, 3199)
- Labels:
- "cell_imbels': DataTree(yx) (17698, 51187), (8549, 25593), (4274, 12766), (2137, 6398), (1668, 3199)
- "cell_imbels': DataTree(yx) (17698, 51187), (8549, 25593), (4274, 12766), (2137, 6398), (1668, 3199)
- "cell_oundaries': DataTree with shape: (4Delayed», 11) (3D points)
- "transcripts": DataTree with shape: (4Delayed», 11) (3D points)
- "cell_circles': GeodataTrame shape: (162254, 21) (3D shapes)
- "cell_circles': GeodataTrame shape: (15254, 2) (3D shapes)
- "micleus_boundaries': GeodataTrame shape: (15628, 1) (2D shapes)
- "tables:
- "tables: AnnOnta (162254, 377)
with coordinate systems:
- "global", with elements:
- he_image (Images), morphology, focus (Images), cell_labels (Labels), nucleus_labels (Labels), transcripts
(Polats), cell_boundaries (Shapes), cell_circles (Shapes), nucleus_boundaries (Shapes)
```

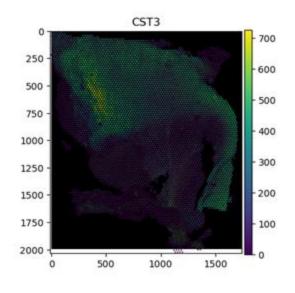
### spatialdata-plot enables static, composable visualization

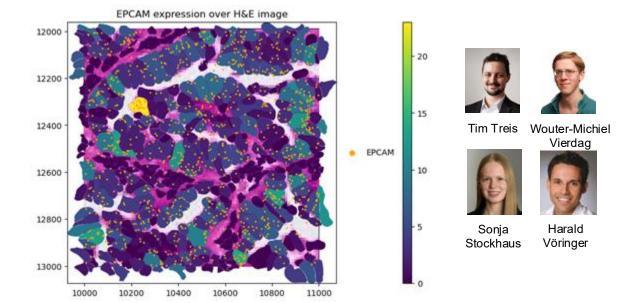
```
import spatialdata_plot

(
    sdata_visium
    .pl.render_images()
    .pl.render_shapes(color="CST3")
    .pl.show("downscaled_hires", title="CST3")
)
```

```
import spatialdata_plot

(
    sdata_xenium.
    .pl.render_images("he_image")
    .pl.render_shapes("cell_boundaries", color="EPCAM")
    .pl.render_points("transcripts", color="feature_name", groups="EPCAM", palette="orange")
    .pl.show(title=f"EPCAM expression over H&E image", coordinate_systems="global", figsize=(10, 5))
)
```



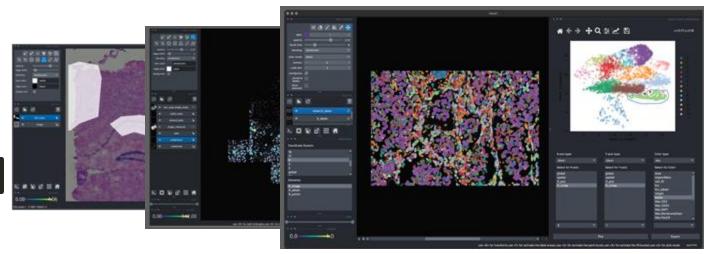


# Napari SpatialData enables interactive data visualization and annotation

## Napari plugin



napari\_spatialdata.Interactive(cosmx\_sdata)





Giovanni Palla



Wouter-Michiel Vierdag

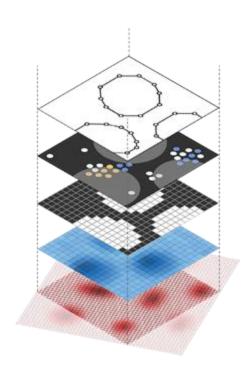


Marcela Toth



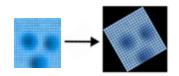
Rahul B. Shrestha

### Generalized, reusable operations are defined for SpatialData objects



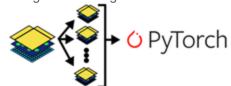
#### **Coordinate transformations**

e.g. rotate an image



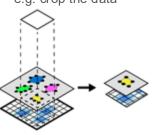
#### **Deep learning interface**

e.g. create image tiles around cells



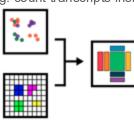
#### **Spatial queries**

e.g. crop the data



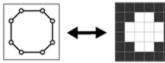
#### **Spatial aggregations**

e.g. count transcripts inside cells



#### Rasterize, vectorize





#### Documentation:

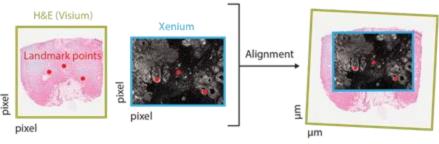
https://spatialdata.scverse.org/en/stable/

Some usage examples of these operations

# Aligning multiple modalities with a landmark-based workflow

#### Before alignment (raw data)

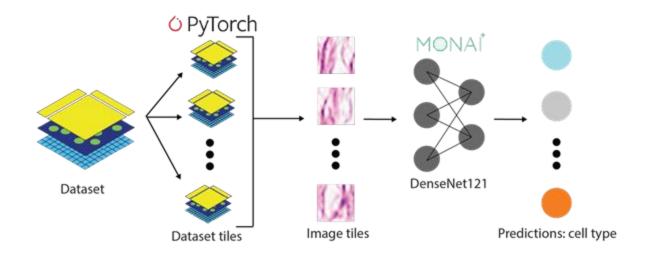




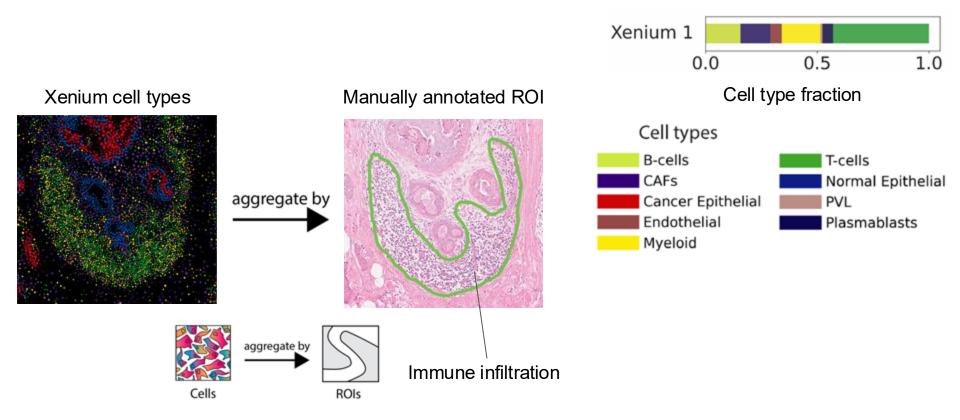
After alignment



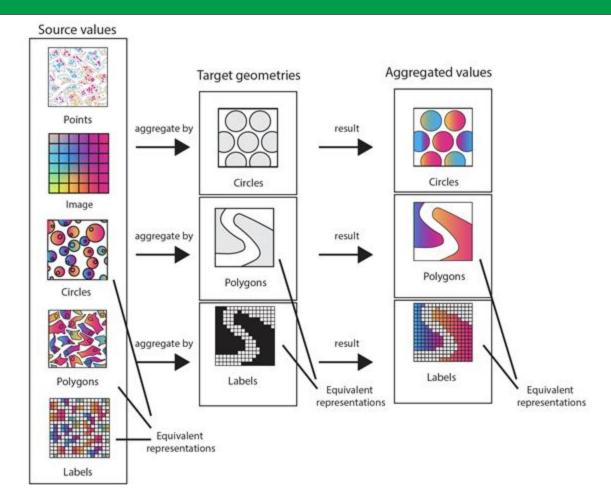
# Creating deep learning datasets from multiple aligned modalities



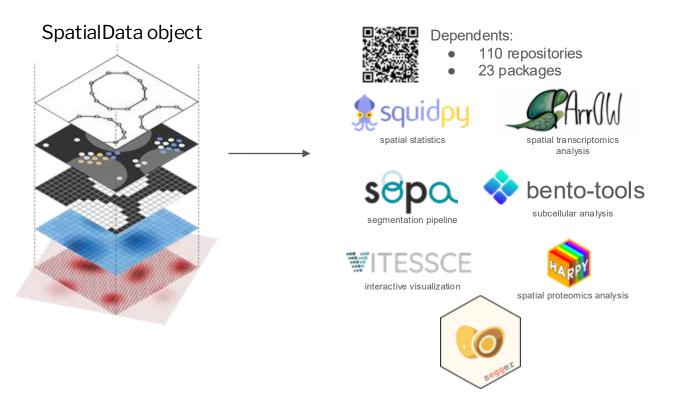
## Aggregation example: computing cell types fractions within regions of interest



## Aggregation is supported for different coordinate systems and representations

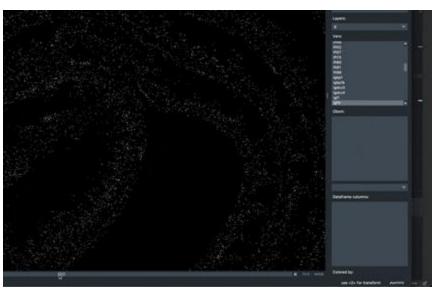


### The community is enabling interoperable analysis of SpatialData objects via a growing ecosystem



### Very performant visualization of 2µm Visium HD bins in napari using rasterize\_bins()





~1000x speed-up for visualizing Visium HD data in napari



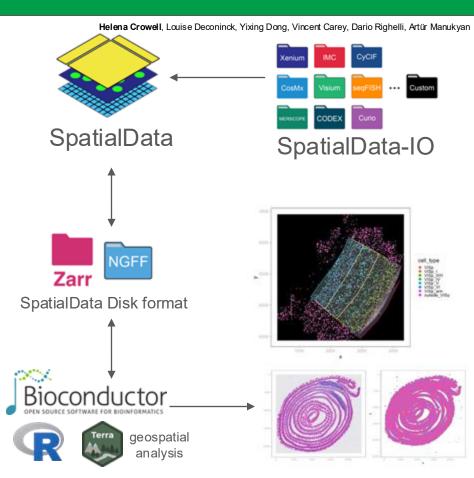
# We are supporting Bioconductor developers building on top of SpatialData Zarr



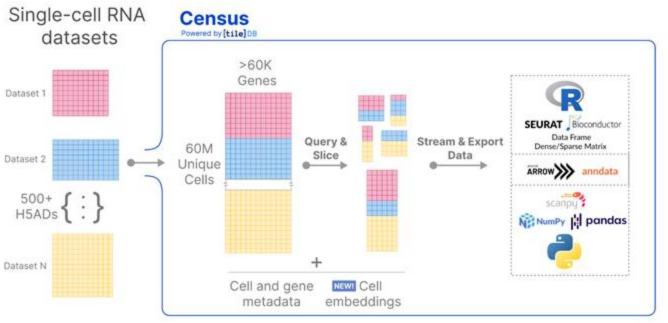
Supported by: Chan
Zuckerberg
Initiative @



SpatialData Hackathon - Nov 2024, Basel (Switzerland) Python & R developers working together



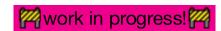
#### The CZI CELLxGENE Census database is expanding to spatial, and now exports to SpatialData





From https://cellxgene.cziscience.com/

# We are preparing a public database of curated spatial omics datasets



# load a SpatialData dataset from the cloud
sdata\_section1\_query = ln.Artifact.filter(
 organisms=organisms.human,
 tissues=tissues.breast,
 description\_\_icontains="Block A Section 1"
).one().load()
sdata\_section1\_query



example notebook (preview)

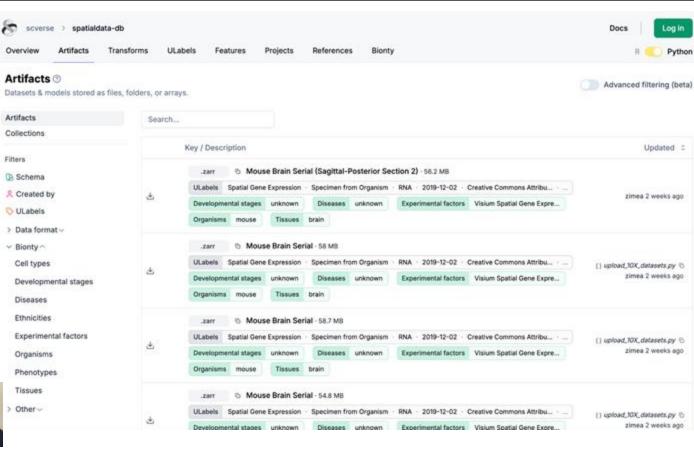


Tim Treis Lea Zimmermann

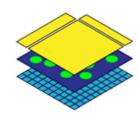
Lukas Heumos



Mark Keller



# Conclusions and acknowledgements



The Spatial Data framework:

- data standard for spatial omics
- spatial operations (aggregation, ROI selection, rasterization, ...)
- current work on robustness, scalability
  - Not an analysis library



Luca Marconato



Kevin Yamauchi



Isaac Virshup



**Tim Treis** 



Michiel Vierdag

Josh Moore























Giovanni

Palla













Sonia Stockhaus Heidari



Marcela Toth

Quentin Blampey

Laurens Lehner

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Lotte **Pollaris** 

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Oliver Stegle

Fabian Theis

Moritz Gerstung

Saka

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Yvan Wolfgang Huber Saeys

Nature Methods, 2024

First authors are underlined

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